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FAILURE OF PROTECTIVE ACTION OF SODIUM AND POTASSIUM IONS
AGAINST HEAT INACTIVATION OF *ESCHERICHIA COLI* L-ASPARAGINASE

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SUMMARY

Two L-asparaginases (L-asparagine amidohydrolase, EC 3.5.1.1) isolated from *Escherichia coli* B were not protected from heat inactivation by sodium (or potassium) ions.

INTRODUCTION

It was shown that certain guinea pigs possessed a thermostable L-asparaginase (L-asparagine amidohydrolase, EC 3.5.1.1) in their sera, while others possessed a thermolabile form of the enzyme, and these two L-asparaginases were stabilized against heat inactivation by Na^+ and K^+ ¹. Roberts *et al.*² and Campbell *et al.*³ demonstrated that *Escherichia coli* B L-asparaginase existed in two forms, one active and the other inactive against animal tumors. These two enzymes differ markedly by several criteria: solubility in ammonium sulfate solution, chromatographic behavior, enzyme activity as a function of pH, as well as the antitumor activity. Are these two *E. coli* L-asparaginases also protected from heat inactivation by Na^+ (or K^+), as well as guinea pig serum L-asparaginase? The present communication reports the effects of Na^+ and K^+ on the heat inactivation of two L-asparaginases isolated from *E. coli* B.

MATERIALS AND METHODS

Enzyme purification

Asparaginase was purified from *E. coli* B by the method of Roberts *et al.*², with some modifications.

Enzyme activity

L-Asparaginase assays were carried out according to the procedures described by Roberts *et al.*². One international unit of activity is defined as that amount of enzyme which catalyzes the formation of 1 μ mole of ammonia per min. Specific activity is expressed as units per mg of protein. The protein content of the samples was determined according to the method of Lowry *et al.*⁴.

RESULTS AND DISCUSSION

Two L-asparaginase components are isolated from *E. coli* and these two enzymes differ markedly in their pH-activity profiles, their solubility, chromatographic behavior, their affinities for L-asparagine, their sensitivities to thermal inactivation, blood clearance and antitumor activity as described in earlier publications^{2,3,5,6}.

Several properties of two peaks of L-asparaginase activity separated by DEAE-cellulose column chromatography were examined, and subsequently were ascertained to be EC-2 and EC-1, respectively. Since the preparations of EC-2 and EC-1 were eluted from the column with a salt gradient and then concentrated, these enzyme preparations contained much salt. In order to remove much of the NaCl in the eluate, these preparations were dialyzed against 0.01 M NaCl. Specific activities of EC-2 and EC-1 were 3.4 I.U. per mg protein and 0.3 I.U. per mg protein, respectively.

Effect of Na⁺ and K⁺ on L-asparaginase EC-2

The activity of EC-2 was slightly (17.5% of initial asparaginase activity) lost after incubation for 30 min at 56 °C, but at 66 °C the enzyme activity was almost entirely (89.3%) destroyed. The thermolabile asparaginase activity in the guinea pig serum sample, which was entirely destroyed at 66 °C for 30 min, was completely (91.8%) stable even after heating in the presence of 0.75 M NaCl¹. Accordingly, the protective effect of Na⁺ (or K⁺) against heat inactivation of EC-2 was examined.

TABLE I

HEAT STABILITY OF THE ACTIVITY OF EC-2 INCUBATED IN THE PRESENCE OF Na⁺ (OR K⁺)
Initial activity taken as 100%.

| Salt | Final concentration (M) | L-Asparaginase activity after incubation at 66 °C for 30 min (% of initial activity) |
|------|-------------------------|--|
| None | — | 10.7 |
| NaCl | 0.05 | 9.4 |
| | 0.25 | 21.8 |
| | 0.75 | 28.8 |
| | 1.0 | 23.2 |
| | 1.25 | 24.8 |
| KCl | 0.05 | 12.8 |
| | 0.25 | 24.0 |
| | 0.75 | 25.5 |
| | 1.0 | 24.8 |

NaCl (or KCl) at different concentrations was mixed with an equal volume of EC-2 preparation and the mixture was heated at 66 °C for 30 min, and then aliquots of the mixture were withdrawn and assayed for L-asparaginase activity. These results are shown in Table I. In the presence of a high concentration of Na⁺, the activity was only slightly stable after heating (0.25 M, 21.8%; 0.75 M, 28.8%; 1.0 M, 23.2%; 1.25 M, 24.8%), whereas in the presence of a low concentration of Na⁺, the activity was not protected from heat inactivation (0.05 M, 9.4%). K⁺ also showed a similar effect on EC-2.

Effect of Na⁺ and K⁺ on L-asparaginase EC-1

After 30 min at 56 °C, 91.7% of the activity of EC-1, which coincided closely with the data reported by Schwartz *et al.*⁵, was lost. Similarly, the activity of EC-2 was inactivated at 66 °C, whereas the inactivating temperature for EC-1 was 10 °C lower. Therefore, the protective effect of Na⁺ (or K⁺) during incubation at 56 °C on EC-1 was investigated. These results are indicated in Table II. The activity of EC-1 was also not protected from heat inactivation by Na⁺ (or K⁺). Thus Na⁺ (or K⁺) showed no significant protective effect on *E. coli* L-asparaginase, in contrast to the

TABLE II

HEAT STABILITY OF THE ACTIVITY OF EC-1 INCUBATED IN THE PRESENCE OF Na⁺ (OR K⁺)
Initial activity taken as 100%.

| Salt | Final concentration (M) | L-Asparaginase activity after incubation at 56 °C for 30 min (% of initial activity) |
|------|-------------------------|--|
| None | — | 8.3 |
| NaCl | 0.05 | 6.8 |
| | 0.25 | 8.7 |
| | 0.75 | 7.4 |
| | 1.0 | 6.7 |
| KCl | 0.05 | 6.8 |
| | 0.25 | 8.0 |
| | 0.75 | 7.3 |
| | 1.0 | 7.8 |

full protective effect of these ions on the activity of guinea pig serum L-asparaginase.

E. coli L-asparaginase (EC-2) and guinea pig serum L-asparaginase differ in their affinities for L-asparagine, molecular weights⁷⁻⁹, the detection of the antitumor activity depending upon the method of testing¹⁰ and blood clearance⁶. Therefore, a remarkable difference in the protective effect of Na⁺ against heat inactivation between *E. coli* L-asparaginase and guinea pig serum L-asparaginase may be related to the difference in these properties of the two enzymes.

REFERENCES

- 1 C. Ryoyama, *Biochim. Biophys. Acta*, 236 (1971) 8.
- 2 J. Roberts, M. D. Prager and N. Bachynsky, *Cancer Res.*, 26 (1966) 2213.
- 3 H. A. Campbell, L. T. Mashburn, E. A. Boyse and L. J. Old, *Biochemistry*, 6 (1967) 721.
- 4 O. H. Lowry, N. J. Rosebrough, A. L. Farr and R. J. Randall, *J. Biol. Chem.*, 193 (1951) 265.
- 5 J. H. Schwartz, J. Y. Reeves and J. D. Broome, *Proc. Natl. Acad. Sci. U.S.*, 56 (1966) 1516.
- 6 E. A. Boyse, L. J. Old, H. A. Campbell and L. T. Mashburn, *J. Exp. Med.*, 125 (1967) 17.
- 7 T. O. Yellin and J. C. Wriston, *Biochemistry*, 5 (1966) 1605.
- 8 J. Kirschbaum, J. C. Wriston and O. T. Ratych, *Biochim. Biophys. Acta*, 194 (1969) 161.
- 9 P. P. K. Ho, E. B. Milikin, J. L. Bobbitt, E. L. Grinnan, P. J. Burck, B. H. Frank, L. V. D. Boeck and R. W. Squires, *J. Biol. Chem.*, 245 (1970) 3708.
- 10 L. T. Mashburn, E. A. Boyse, H. A. Campbell and L. J. Old, *Proc. Soc. Exp. Biol. Med.*, 124 (1967) 568.